Data Visualisation with ggplot2

Communicating your data

Daniela Palleschi

2024-06-03

Table of contents

Learning objectives	2
Load packages and data Summary of first-fixation times	2
Plotting reading times	3
Plots with ggplot2	3
An example: histogram	3
Start layering	4
Add labels	5
Add	6
Add condition	7
Customisation	8
theme_bw()	8
theme_minimal()	9
Distributions	10
Density plots	11
	12
facet_grid()	13
	14
Scatterplots	16
Scatterplots	17
Bar plot	18
•	20
	21

Grouped bar plots	22
	40
Extra exercise	
Summary statistics	25
Boxplots	25
Boxplot explained	27
Boxplots	
Grouped boxplots	28
Interaction plots	29
Question: Binomial data	31
Saving our plots	31
ggsave()	32
saveRDS()	32
readRDS()	33

Learning objectives

- visualise variable distributions
- visualise summary statistics
- save figures as rds or as figures

Load packages and data

Summary of first-fixation times

• this code is from the previous topic

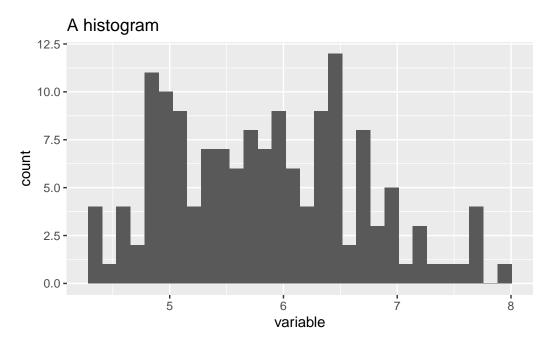
Plotting reading times

- reading times are (usually) continuous variables
 - as are e.g., reaction times
- they are truncated at 0, meaning they cannot have negative values
 - because of this, they tend to have a skewed distribution

Plots with ggplot2

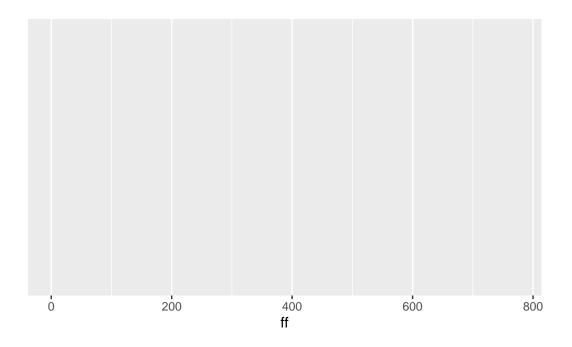
- ggplot2 is part of the tidyverse (like dplyr)
 - uses a layered grammar of graphics
 - i.e., we build layers

An example: histogram



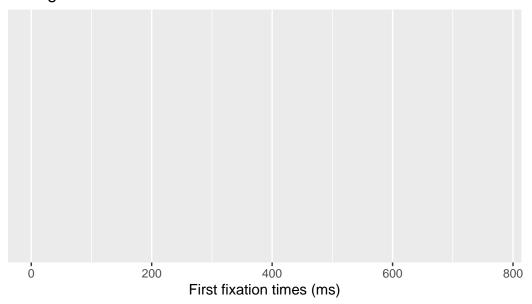
Start layering

```
df_lifetime |> ggplot(aes(ff)) # aes = 'aesthetic'
```



Add labels

Histogram of first fixation times



Add

Histogram of first fixataion times

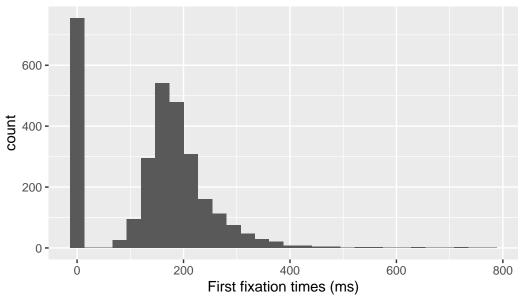
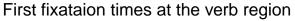


Figure 1: Distribution of first fixation times at the verb region (raw milliseconds)

Add condition



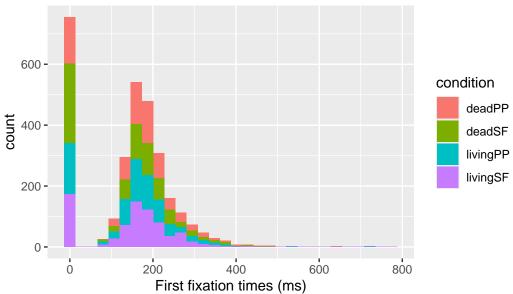


Figure 2: Distribution of first fixation times at the verb region (raw milliseconds)

The colour here is STACKED!! i.e., not layered. Notice the distribution doesn't change from all grey to coloured

Customisation

- we can add arguments to our geoms
 - e.g., transparency: alpha = takes a value between 0 to 1
- we can use theme() to customise font sizes, legend placement, etc.
- tehre are also popular preset themes, such as theme_bw() and theme_minimal()

theme_bw()

Histogram of first fixataion times

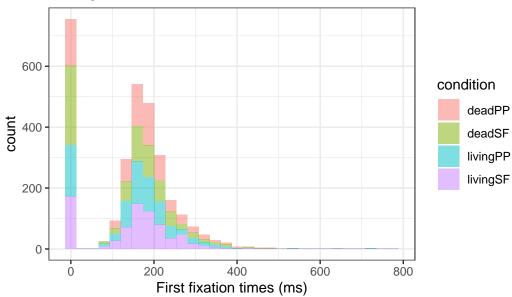


Figure 3: Distribution of first fixation times at the verb region (raw milliseconds).

theme_minimal()

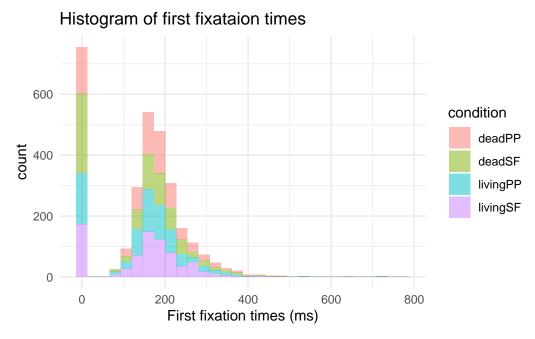
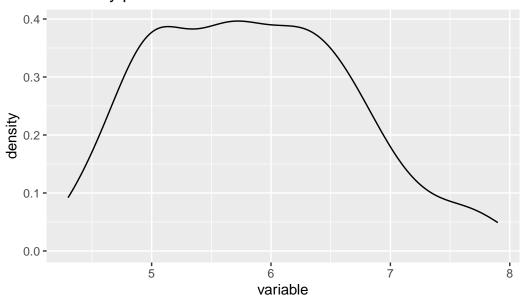


Figure 4: Distribution of first fixation times at the verb region (raw milliseconds).

Distributions

- show the distribution of observations
 - so we can see where the data are clustered
 - and eyeball the shape of the distribution
- we already saw the histogram, which shows the number of observations per variable value
- density plots are another useful plot for visualising distributions

A density plot



Density plots

- below I just replaced geom_histogram() with geom_density()
 - I also filtered the data to include only values of ff above 0
- what is plotted along the y-axis? how does this differ from a histogram?

Histogram of first fixataion times

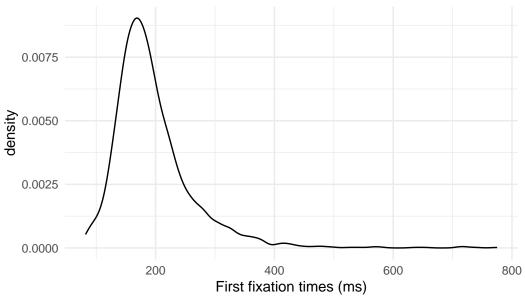


Figure 5: Distribution of first fixation times at the verb region (raw milliseconds).

Grouped density plots

- just like with histograms, we can look at the density plots of different subsets of the data with aes(fill =)
 - like region

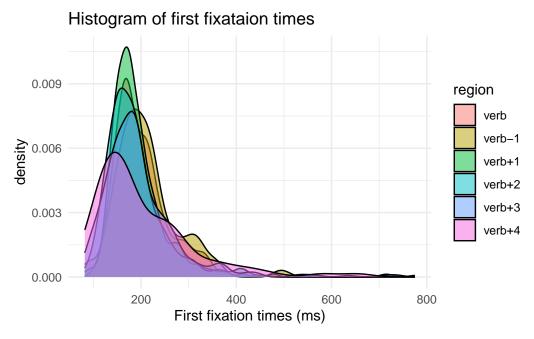


Figure 6: Distribution of first fixation times at the verb region (raw milliseconds).

facet_grid()

• there are a lot of overlapping density curves, let's try to separate them with facet_grid(x~y)

Density plot of first fixataion times by region

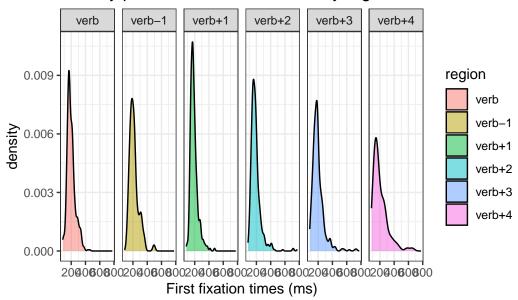


Figure 7: Distribution of first fixation times at the verb region (raw milliseconds).

• how would you describe the density plots of the different regions?

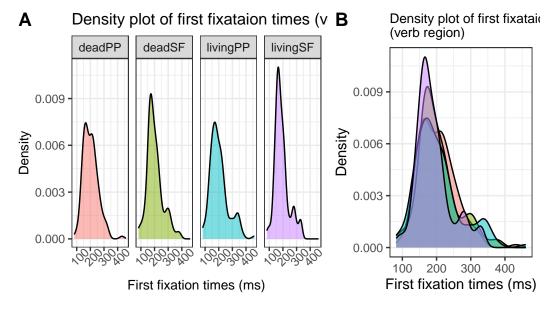
re-ordering factors

- by default, factors will be ordered alphabetically
 - but we don't always want that
 - here, verb-1 should be before verb

verb-1 verb verb+1 verb+2 verb+3 verb+4 559 559 559 559 559 182

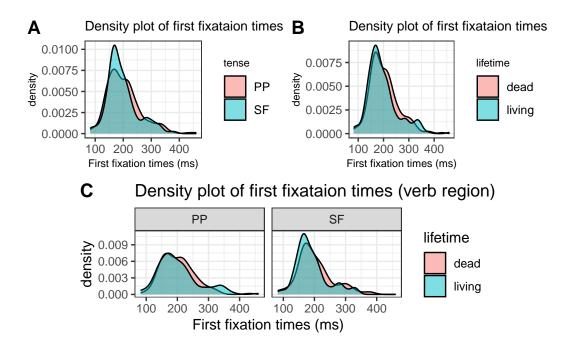
Exercise

- 1. create a density plot with the fill colour set to condition, but:
- subset the data to only include the verb region
- you can decide if you want to use facets or to have the density curves overlayed
- your plot should look something like A or B:



Extra exercise

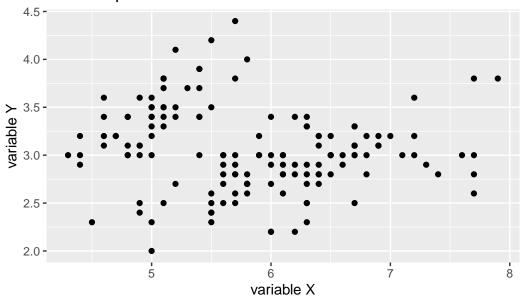
2. Can you produce these plots?



Scatterplots

- histograms and density plots plot a **single variable** along the x-axis
 - in most other plots the dependent (measure) variable is plotted along the y-axis by convention
- scatterplots plot the relationship between two variables

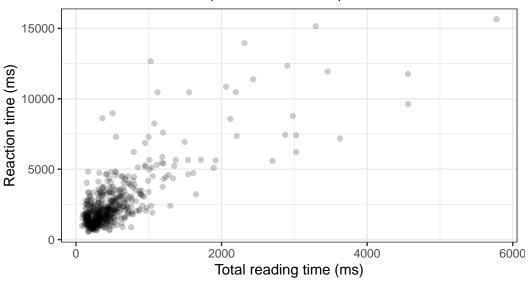
A scatterplot



Scatterplots

- the figure below plots total reading times (verb region) to the verb region (x-axis) and reaction times to the critical sentence (y-axis)
 - what does each point represent?
 - how would you describe the relationship between the two variables?

Scatter plot of total reading times (verb region) and reaction times (critical sentence)



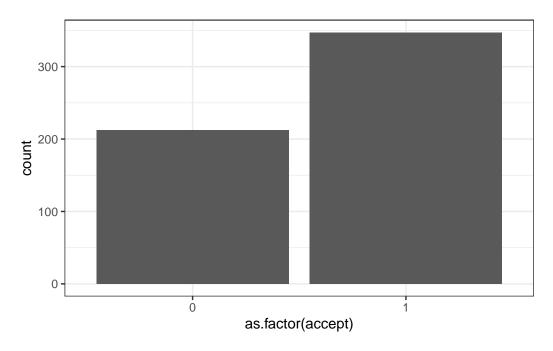
Exercise

- 1. Generate a scatterplot of total reading times and reaction times, with:
 - colour and shape set to condition
 - tip: these both belong in aes()
- 2. What information does this plot suggest?

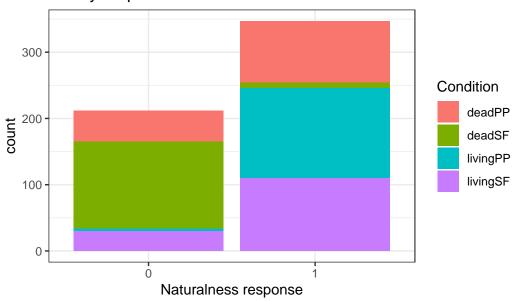
Bar plot

- show the distribution of categorical factor levels
 - i.e., the frequency of observations per level
- be sure to read in accept as a factor!

```
df_lifetime |>
  distinct(px,trial,.keep_all=T) |>
  ggplot(aes(x = as.factor(accept) )) +
  geom_bar() +
  theme_bw()
```

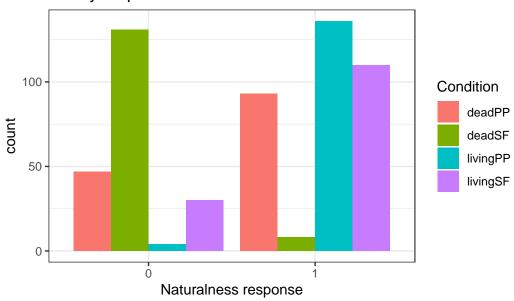


Binary responses



Grouped bar plots

Binary responses

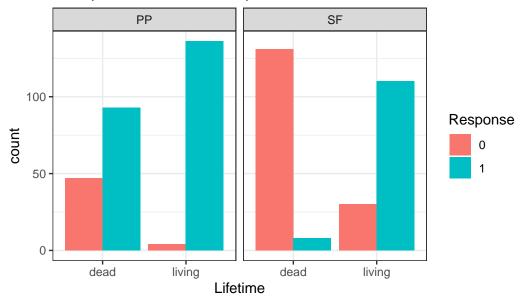


Exercise

- 1. Generate a grouped bar plot (i.e., dodge) with:
 - a facet grid for tense
 - plots lifetime on the x-axis
 - and fills the bars based on accept
 - change the labels accordingly
 - customise as you like

Grouped bar plots

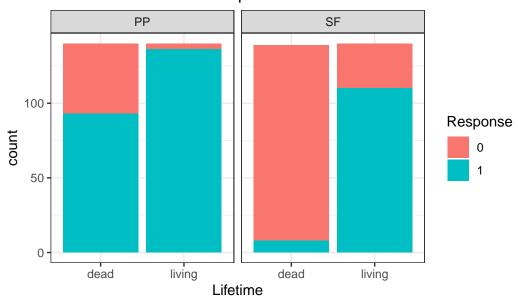
Grouped and faceted barplot



Stacked bar plots

```
geom_bar(position = "stack") +
theme_bw()
```

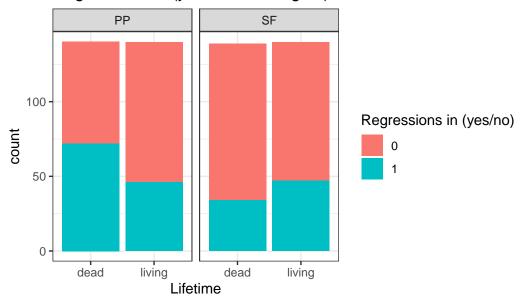
Stacked and faceted barplot



Exercise

- 1. Choose the barplot you like best for binary data
- 2. Reproduce that barplot, but with ${\tt reg_in}$ at the ${\tt verb1}$ region

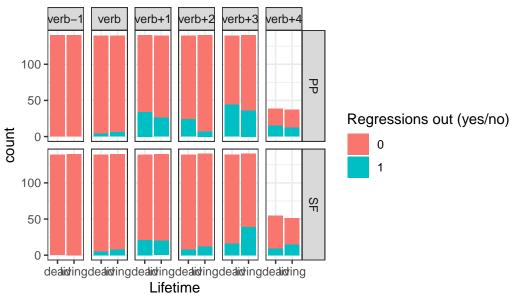
Regressions in (yes/no, verb region)



Extra exercise

- 1. Create another bar plot, but for reg_out for all sentence regions
- Use facet_grid()
- to have facets by region (columns) and by tense (in 2 rows)





Summary statistics

- measures of location: mean, median, mode
- measures of spread: (interquartile) range, standard deviation

Boxplots

- boxplots provide information about the distribution of a *continuous* variable
 - but includes information like *median* (dark line) and *quartiles* (box and whiskers)
 - and outliers (dots)
- like scatterplots, require x and y variables
 - but one of them needs to be **categorical**

```
iris |>
  ggplot(aes(x = Species, y = Sepal.Length)) +
  labs(title = "A scatterplot",
        x = "Categorical variable",
        y = "Continuous variable") +
  geom_boxplot()
```

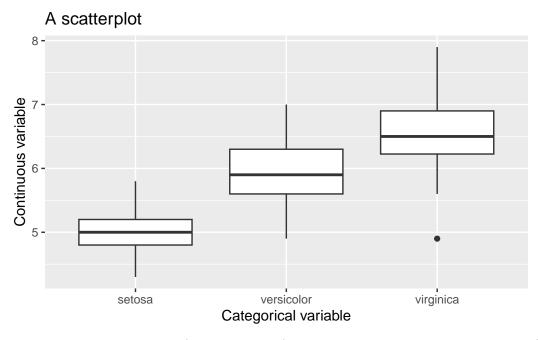


Figure 8: A scatterplot. Median (50th percentile): thick black lines; interquartile range (IQR; 25th and 75th percentile): box limits; minimum (0th percentile) and maximum (100th percentile) excluding outliers: : whiskers; outliers: points

Boxplot explained

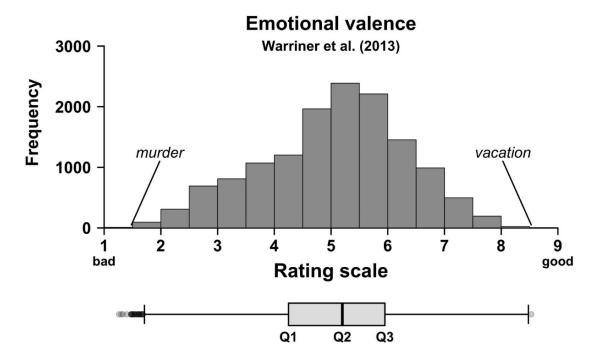


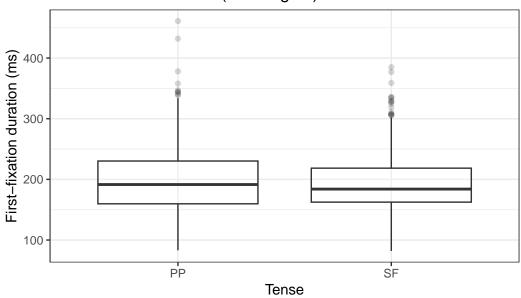
Figure 3.4. A histogram of the emotional valence rating data

Figure 9: Image source: Winter (2019) (all rights reserved)

Boxplots

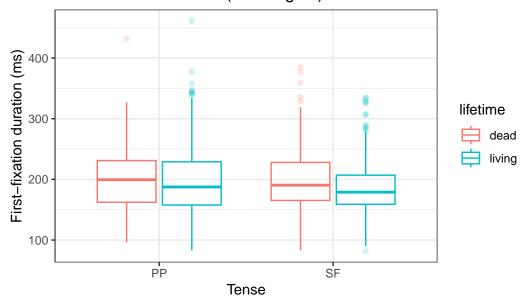
• let's change our scatterplot to a boxplot

First-fixation duration (verb region)



Grouped boxplots

First-fixation duration (verb region)



Exercise

- 1. Create a group boxplot (x = tense, fill = lifetime) for
- first-pass reading time (verb region)
- regression path duration (verb region)
- total reading time (verb region)
- reaction times (use the distinct() verb to have a single observation per participant and per trial)

Interaction plots

- common for **factorial** designs, i.e., comparing *categorical* predictors
- there are 2 ways of producing them:
 - with your data frame and stat_summary()
 - or with a summary table and ggplot geoms geom_point(), geom_errorbar(), and geom_line()
- we'll need our summary table to plot an interaction plot

condition	lifetime	tense	N	mean.ff	sd	se	ci	lower.ci	upper.ci
deadPP	dead	PP	140	198.9	57.9	4.9	9.7	189.2	208.6
deadSF	dead	SF	139	194.6	67.9	5.8	11.4	183.2	205.9

condition	lifetime	tense	N	mean.ff	sd	se	ci	lower.ci	upper.ci
livingPP	living	PP	140	194.2	77.3	6.5	12.9	181.3	207.1
livingSF	living	SF	140	186.0	57.6	4.9	9.6	176.4	195.6

```
library(patchwork)
df_lifetime |>
  filter(region == "verb") |>
  ggplot(aes(x = lifetime, y = ff,
                     shape = tense,
                     group = tense,
                     color = tense)) +
  labs(title="Interaction plot (`stat_summary()`)",
       x = "Lifetime",
       y = "First fix (ms)",
       shape = "Tense", group = "Tense", color = "Tense", linetype = "Tense") +
  stat_summary(fun = "mean", geom = "point", size = 3, position = position_dodge(0.2)) +
  stat_summary(fun = "mean", geom = "line", position = position_dodge(0.2), aes(linetype=tens
  stat_summary(fun.data = "mean_cl_normal", geom = "errorbar", width = .2
               , position = position_dodge(0.2)) +
  theme_bw() +
summary_ff |>
  ggplot(aes(x = lifetime, y = mean.ff,
                     shape = tense,
                     group = tense,
                     color = tense)) +
  labs(title="Interaction plot (geoms)",
       x = "Lifetime",
       y = "First fix (ms)",
       shape = "Tense", group = "Tense", color = "Tense", linetype = "Tense") +
  geom_point(size = 3,
                position = position_dodge(0.2)) +
  geom_line(aes(linetype=tense), position = position_dodge(0.2)) +
  geom_errorbar(aes(ymin = mean.ff - ci,
                    ymax = mean.ff + ci),
                width = .2,
                position = position_dodge(0.2)) +
  theme_bw() +
  plot_annotation(tag_levels = "A") +
  plot_layout(guides = "collect") &
```

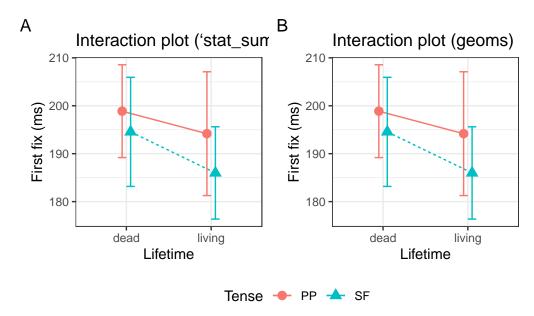


Figure 10: Identical interaction plots produced two ways: feeding a dataset into ggplot() and using stat_summary() (A) or feeding a summary table into ggplot() and using geoms (B)

Question: Binomial data

- binomial data are those with 2 categories, for example
 - present, absent
 - yes, no
- in our dataset, each trial ended with a binary naturalness judgement task
 - how might we plot such data?

Saving our plots

- we can save our plots two ways:
 - 1. as image files (e.g., JPEG, PNG, SVG, etc.): when writing in Word, LaTeX, etc.
 - 2. as a single R object (Rds: R data structure): when writing in Rmarkdown/Quarto

ggsave()

- the ggsave() function is useful for saving ggplot objects
 - we first have to save one of our figures as an object
 - I usually save ggplot objects with the prefix fig_ (short for figure)
- make sure you also have a useful place to store these figures
 - e.g., a folder called figures

```
fig_lifetime_ff <-
summary_ff |>
  ggplot(aes(x = lifetime, y = mean.ff,
                     shape = tense,
                     group = tense,
                     color = tense)) +
  labs(title="Mean first-fixation times (verb region) with 95% CIs",
      x = "Lifetime",
       y = "First fix (ms)",
       shape = "Tense", group = "Tense", color = "Tense", linetype = "Tense") +
  geom_point(size = 3,
                position = position_dodge(0.2)) +
  geom_line(aes(linetype=tense), position = position_dodge(0.2)) +
  geom_errorbar(aes(ymin = mean.ff - ci,
                    ymax = mean.ff + ci),
                width = .2,
                position = position_dodge(0.2)) +
  theme_bw()
```

```
ggsave(fig_lifetime_ff, filename = here("figures", "fig_lifetime_ff.png"))
```

- ggsave() has lots of arguments to control width, height, resolution, etc.
 - to see more, run ?ggsave in the Console
- you can also save as JPG/JPEG, SVG, even PDF by just changing the filename extension

saveRDS()

- we can also save the figure as R code
 - which means we can control the width, height, resolution, etc. later on when we load it in

- useful if you'll be writing up your results in R markdown or Quarto

```
saveRDS(fig_lifetime_ff, file = here("figures", "fig_lifetime_ff.rds"))
```

readRDS()

- you can't click on the file to view the figure because it's R code
 - you'd need to load the data into R again

```
fig_lifetime_ff <- readRDS(here("figures", "fig_lifetime_ff.rds"))</pre>
```

Naming files and saving code

You'll notice I saved the PNG and RDS files using the same name that the I used for the figure in my script. This is an important point: I want to be able to traceback my figures from the code so I can easily track them. It also helps encourage informative object and file names.

Of course, saving the code used to save the files in our scripts is also useful because we can easily adjust the saved files (e.g., change figure width or height)

References

Nordmann, E., & DeBruine, L. (2022). Applied data skills. Zenodo. https://doi.org/10.5281/zenodo.6365078

Nordmann, E., McAleer, P., Toivo, W., Paterson, H., & DeBruine, L. M. (2022). Data Visualization Using R for Researchers Who Do Not Use R. Advances in Methods and Practices in Psychological Science, 5(2), 251524592210746. https://doi.org/10.1177/25152459221074654

Wickham, H., Çetinkaya-Rundel, M., & Grolemund, G. (2023). R for Data Science (2nd ed.). Winter, B. (2019). Statistics for Linguists: An Introduction Using R. In Statistics for Linguists: An Introduction Using R. Routledge. https://doi.org/10.4324/9781315165547